

Fast, Regional 3D Hybrid (1D-Hadamard 2D-Rosette) Proton MRSI in The Human Brain

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Introduction

Fast 3D ¹H-MRSI methods cover large regions to full brain, at $\lt 1\text{ cm}^3$ spatial resolution, in $\lt 20$ minute (1). However, sometimes, only regional coverage is desired. e.g., in lower brain regions, near the air-filled sinuses, where 1-2 ppm/cm local \mathbf{B}_0 variations are encountered (2), undermining water suppression and SNR. Their removal requires shim orders not found on clinical scanners (3), but they can be mitigated by: (a) shimming only the structure(s) sought (3); (b) small voxels to sample less \mathbf{B}_0 variations (4); and (c) thin VOI in the susceptible direction (5). The problem with these is that if CSI is used to localize in the thin direction, its spatial response (PSF), will: (a) contaminate the VOI with extraneous signals from the imperfect slice profile; (b) bleed-in signal between slices; and (c) bleed-out up to 13%, lowering the SNR (6). All three, we show, can be addressed with cascaded transverse Hadamard spectroscopic imaging (cT-HSI) for localization along the slice-selection direction (7). Unlike CSI, cT-HSI PSF can be improved to a nearly ideal "boxcar" with high BW optimized RF pulses, minimizing both bleeds, maximizing the VOI SNR (8). Here we demonstrate these advantages by comparing $\times 2$ cT-HSI with $\times 8$ CSI, both with in-plane 2D 20×20 rosette-spectroscopic-imaging (RSI) (9), for fast, $\lt 10$ minute, 3D ¹H-MRSI in a phantom comprising partitions of different metabolites, to identify bleeds and localize their sources and in the human hippocampus.

Methods

Human and phantom data were acquired on a 3 T MR scanner and 20-channel head-coil. Two 3D ¹H-MRSI sequences were compared. One defined the 2 cm thick VOI (to avoid local susceptibility inhomogeneities too strong or varying to shim, e.g., in the temporal lobes) with a 3 ms, 18 μT peak \mathbf{B}_{1+} Shinnar-Le Roux (SLR) 90° pulse, using $\times 8$ CSI to localize in that direction. The other uses a cascade of two of the same SLR 90°s for $\times 2$ cT-HSI encoding under $\times 2$ fold stronger gradients. For both, the other two directions were localized with 2D 20×20 rosette spectroscopic imaging (RSI).

Results

cT-HSI is shown both (a) in simulations; (b) a structured phantom; and (c) *in vivo* in the human temporal lobes, to dramatically reduce the signal bleed within the VOI slices from the $\sim 26\%$ in CSI to $\lt 1\%$ in simulations; and from $\sim 50\%$ down to 5-8% in a phantom. The cT-HSI SNR is improved by 10-15% compared with the $\times 8$ CSI, reflecting the former's commensurate reductions in signal bleed out.

Discussion

Since in MRSI spatial and spectral localizations are the *raison d'être*, it is shown that CSI spatial and spectral localization shortcomings, can be dramatically reduced to negligible fractions, using high BW cT-HSI RF pulses (7). This advantage, however, comes at the costs of increased SAR from the RF pulses, progressively larger T_2 weighting for earlier pulses in long, $\gt 4$, cascades and loss of acceleration in the HSI direction(s). Nevertheless, hybrids of cT-HSI in the VOI narrow direction(s) with RSI in-plane, e.g., as shown here, can be conceived to take advantage of each method's advantages, while minimizing the impact of their weaknesses.

Conclusion

Compared with CSI, cT-HSI hybrid with in-plane RSI, facilitates fast 3D multivoxel encoding $20\times 20\times 2$ voxels at sub 1 cm^3 spatial resolution over the bilateral human hippocampus, in under 5 minutes at negligible spectral and spatial contamination and $\sim 15\%$ SNR gains compared with the customary CSI.

References

1. Lecocq *et al.*. J Magn Reson Imaging 2015;42(2):280-289.
2. Li S, *et al.* Magn Reson Med 1996;36(5):705-714.
3. Pan JW *et al.* Magn Reson Med 2012;68(4):1007-1017.
4. Li BS, *et al.* Magn Reson Med 2001;46(6):1049-1053.
5. Schirda CV *et al.* Magn Reson Med 2018;79(5):2470-2480.
6. Maudsley AA. Journal of Magnetic Resonance 1986;68:363-366.
7. Goelman G, *et al.* Magn Reson Med 2007;58(1):167-173.
8. Cohen O, *et al.* Magn Reson Med 2014;72(4):923-933.
9. Schirda CV *et al.* Magn Reson Med 2015.

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